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## PREPARATION OF OPTICALLY ACTIVE 4-ALLYLAZETIDIN-2-ONES. AN ACCESS TO CARBACEPHAMS

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Abstract: Transesterification of 4-allyl-1-hydroxymethylazetidin-2-ones using vinyl acetate in the presence of the lipase from  $Pseudomonas\ cepacia\$ led to the corresponding (R)-acetates and the remaining (S)-alcohols in high yields and excellent ee's (E:32-71). Subsequent reaction with BF<sub>3</sub>-Et<sub>2</sub>O can lead to the carbacepham framework.

A major problem, which has appeared in the last years, is the bacterial resistance to antibiotics.<sup>1</sup> One way to overcome this difficulty is the design of new products which cannot be obtained by fermentation. Promising results have been recently obtained with carbacephems,<sup>2</sup> which show a microbiological activity comparable to the cephalosporin equivalents, associated with an enhanced physiological half-life.<sup>3</sup> Numerous carbacephem synthetic approaches have been explored,<sup>4</sup> but only a few led to optically active compounds.<sup>5</sup> Lorabid<sup>®</sup> is the first antibiotic of this new family under clinical development.<sup>6</sup>

We recently reported the enzymatic resolution of γ-butyrolactams by a non-destructive method using a labile N-hydroxymethyl group. We decided to test this method 8 for the preparation of optically active 4-allyl-β-lactams, which could be used as 9 carbacephem precursors.

$$\stackrel{\stackrel{H}{\longrightarrow}}{\underset{O}{\longrightarrow}} \stackrel{\stackrel{H}{\longrightarrow}}{\underset{R^{"}}{\longrightarrow}} \stackrel{\stackrel{H}{\longrightarrow}}{\underset{O}{\longrightarrow}} \stackrel{\stackrel{H}{\longrightarrow}}{\underset{R^{"}}{\longrightarrow}} \stackrel{\stackrel{H}{\longrightarrow}}{\underset$$

The 4-allyl-1-hydroxymethylazetidin-2-ones 1a-1e used in this study have been prepared in two steps from 4-acetoxyazetidin-2-one by reaction first with the corresponding allylsilanes, <sup>10</sup> then formaldehyde <sup>7</sup> (50-80% overall yields). The products were fully characterised by <sup>1</sup>H, and <sup>13</sup>C NMR, and IR spectroscopies.

$$OAC \longrightarrow SiMe_3 \longrightarrow R \longrightarrow R$$

$$O \longrightarrow N \longrightarrow R$$

$$O \longrightarrow N \longrightarrow OH$$

The enzymatic resolutions of these alcohols were conducted in *t*-butyl methyl ether in presence of vinyl acetate using the lipase from *Pseudomonas cepacia* supported on Hyflo Super Cel<sup>®</sup> <sup>7</sup>. Results of the transesterifications are reported in Table 1.

Table 1: Transesterification of alcohols 1a-1e, and chloroacetate 3e

a) Calculated as reported in reference 11. b) Measured in CH<sub>2</sub>Cl<sub>2</sub> (c = 0.5). c) In THF (c = 1).

The overall yields of the transesterification were  $\geq$  95%. Except for alcohol 1e, excellent results were obtained for these transesterifications ( $E \geq 36$ ). In the case of the phenyl substituent the result has been improved using the chloroacetate 3e, prepared by reaction of alcohol 1e with chloroacetic anhydride (CH<sub>2</sub>Cl<sub>2</sub>, 1 eq. DMAP, r.t., 85% yield). In *t*-butyl methyl ether in the presence of the lipase from *Pseudomonas cepacia* and n-propanol the transesterification led to alcohol 1e and the enriched starting chloroacetate (E = 32).

The enantiomeric excesses of the products were measured on the corresponding acetates by  $^{1}H$  NMR spectroscopy in the presence of Eu(hfc)<sub>3</sub>. The absolute configurations of these optically active compounds were determined by chemical correlation with (S)-4-allylazetidin-2-one  $^{12}$  as reported in Scheme 1. We deduced from these results that in all cases examined the enzyme substrate were the (R)-alcohols. With the chloroacetate 3e the (R)-enantiomer was also the enzyme substrate and gave obviously the (R)-alcohol 1e.

Scheme 1: Determination of the Absolute Configuration of  $\beta$ -Lactams 2a - 2e

<sup>a</sup> NH<sub>4</sub>OH, MeOH (50-60%). <sup>b</sup>O<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, -78° C then PPh<sub>3</sub> (65-70%). <sup>c</sup>(COCl)<sub>2</sub>, DMSO, NEt<sub>3</sub> (50%). <sup>d</sup>PhLi, CH<sub>2</sub>Cl<sub>2</sub> (12%). <sup>e</sup>LiOH, H<sub>2</sub>O<sub>2</sub>, H<sub>2</sub>O, THF (50%). <sup>f</sup>tBuOOH, Pd(CF<sub>3</sub>COO)<sub>2</sub> (20%) <sup>g</sup>Ac<sub>2</sub>O, NEt<sub>3</sub>, DMAP, CH<sub>2</sub>Cl<sub>2</sub> (70%). <sup>h</sup>Pd(DBA)<sub>2</sub>, PPh<sub>3</sub>, NaBH<sub>3</sub>CN (55-65%). <sup>i</sup> mCPBA, CH<sub>2</sub>Cl<sub>2</sub>, -30° C (80%). j (CH<sub>2</sub>O)<sub>n</sub>, K<sub>2</sub>CO<sub>3</sub> (70%).

Different Lewis acids such as TiCl<sub>4</sub>, SnCl<sub>4</sub>, AlCl<sub>3</sub> were then tested in the subsequent cyclization reactions to give the carbacepham ring system. The allyl derivative 1a led to the desirated bicyclic compound 4 when treated with SnCl<sub>4</sub> (83% yield). With the substrates 1b-1e and 2b-2e better results were obtained with boron trifluoride etherate. The acetate 2b led to a mixture of products (70% yield), which could not be separated by liquid chromatography.

A much more selective cyclization was observed with the sulfur lactam 2d. In ether in the presence of BF<sub>3</sub>.Et<sub>2</sub>O a single compound 5 of unknown stereochemistry was obtained (75% yield). In the same way cyclization of the chloroacetate (S)-3e gave only 6 (77% yield;  $[\alpha]_D + 14.2 \text{ c} = 0.6 \text{ (THF)}$ ; ee 33%). All these compounds were easily characterised by spectroscopic methods.

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In summary, we have developed a new approach to the carbacepham framework. Extension of this method to the preparation of optically active 1-dethiacephalosporins is under investigation.

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